This article was downloaded by: On: *18 January 2011* Access details: *Access Details: Free Access* Publisher *Taylor & Francis* Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



To cite this Article Blomqvist, S. , Hjellström, K. and Sjösten, A.(1993) 'Interference from Arsenate, Fluoride and Silicate When Determining Phosphate in Water by the Phosphoantimonylmol Ybdenum Blue Method', International Journal of Environmental Analytical Chemistry, 54: 1, 31 - 43**To link to this Article: DOI:** 10.1080/03067319308044425

URL: http://dx.doi.org/10.1080/03067319308044425

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

INTERFERENCE FROM ARSENATE, FLUORIDE AND SILICATE WHEN DETERMINING PHOSPHATE IN WATER BY THE PHOSPHOANTIMONYLMOLYBDENUM BLUE METHOD

S. BLOMQVIST^{1,2,*}, K. HJELLSTRÖM^{3,4} and A. SJÖSTEN¹

¹ Department of Systems Ecology, Section Marine Ecology, ² Department of Geology and Geochemistry, ³ Department of Physical, Inorganic and Structural Chemistry, Stockholm University, S-106 91 Stockholm, Sweden

(Received, 11 February 1993; in final form, 15 April 1993)

In the range of 50–500 μ g PO₄-P L⁻¹, analytical procedures according to Murphy and Riley, ¹ Koroleff² and Parsons *et al.*³ all resulted in complete colour development of the phosphoantimonylmolybdenum blue complex within 5 min after reagent addition. Reducing the reaction temperature from +25° to +15°C slowed the formation rate of the blue complex, but not enough to be of practical importance. Arsenate (0–1.0 mg AsO₄-As L⁻¹), fluoride (0–200 mg F L⁻¹) and silicate (0–50 mg SiO₄-Si L⁻¹) affect the phosphate determination in different ways. Arsenate forms a blue complex with molybdate, but with a slower reaction rate than for the phosphate complexation. At low arsenate concentrations this interference can be reduced by timing the photometric recording according to the phosphoantimonylmolybdenum formation rate. High concentrations of fluoride slow the colour development of the phosphoatimonylmolybdenum blue complex. Silicate may affect the analytical result in two ways. Alone, after prolonged standing, a slighly increased light absorption results from a blue complex formed with molybdate, but solute to fluoride. High fluoride concentrations counteract the development of the phosphoatimonylmolybdenum blue complex. The interfering effects of arsenate, fluoride and silicate can allo reduce the inhibitory effect of results from a blue complex formed with molybdate, but solute to the blue complex between silicate and molybdate. The interfering effects of arsenate, fluoride and silicate can all be reduced by dilution of the sample prior to reagent addition.

KEY WORDS: Phosphate, interference, arsenate, fluoride, silicate, molybdenum blue, phosphoantimonylmolybdenum.

INTRODUCTION

More than 100 years ago, M. F. Osmond⁴ proposed using the blue complexation between

Downloaded At: 14:42 18 January 2011

^{*}Author to whom correspondence should be addressed.

⁴ Present address: Euroc Research AB, Box 104, S-620 30 Slite, Sweden

phosphoric acid and molybdate as a colorimetric analytical tool. Various modifications in the analytical procedure have since been proposed, including adaptations to water samples (see reviews⁵⁻¹¹). The basic chemical principle of the method is formation of a heteropoly phosphate-molybdate acid which, upon reduction, produces a blue colour, the intensity of which is proportional to the amount of orthophosphate ion incorporated into complex form. It has become the standard method for determination of phosphate concentration in water. The modifications most frequently used today are based on addition of ascorbic acid as a reducing agent in the presence of trivalent antimony, as suggested by Murphy and Riley.¹

However, further evaluation of the Murphy and Riley¹ procedure of assaying phosphate seems warranted. In fact, only a few papers have reported rate data on the colour development of phosphoantimonylmolybdenum blue. Jones¹² presented rate measurements of a seawater sample to which $31 \mu g PO_4$ -P L⁻¹ were added, To and Randall¹³ reported measurements in pure water at one phosphate concentration (250 $\mu g PO_4$ -P L⁻¹) and two temperatures (+5° and +22°C), while Pai *et al.*¹⁴ measured at one phosphate concentration (155 $\mu g PO_4$ -P L⁻¹) and four temperatures (+20°, +35°, +55° and +70°C) in deionized distilled water. The recommended time from addition of reagents to the photometric recording, and the time interval within which measurements ought to be made, varies in commonly used handbooks and manuals.^{2-3,15-19} Moreover, many components found in water samples have been reported to interfere with the analysis, potentially biasing phosphate readings. Unfortunately, such reports often give insufficient analytical detail, making it difficult to recommend precautions in order to minimize interference effects. This applies to arsenate, fluoride and silicate, despite early attention to possible biasing interference.^{4,20-23}

We report here (1) data on the reaction rate of phosphoantimonylmolybdenum complexation, involving a comparison of three commonly employed modifications¹⁻³ and an evaluation of the effect of room temperature variation, and (2) some experiments dealing with interference from arsenate, fluoride and silicate.

EXPERIMENTAL

Chemicals. We used doubly distilled or deionized, reagent-grade water and chemicals of p.a. quality (E. Merck, Darmstadt, F.R.G.), i.e. ammonium heptamolybdate tetrahydrate, $(NH_4)_6Mo_7O_{24}\cdot 4H_2O$, L(+) ascorbic acid, $C_6H_8O_6$, potassium antimonyl tartrate, $K(SbO)C_4$ $H_4 O_6 \cdot \frac{1}{2} H_2 O$, potassium dihydrogen phosphate, $KH_2 PO_4$, sodium fluoride, NaF, and sulphuric acid, H_2SO_4 . Sodium fluorosilicate Na₂SiF₆ (Riedel-de Haën AG, Seelze, F.R.G.) and calcium orthoarsenate Ca₃(AsO₄)₂ were of purified quality.

Apparatus. Time-course measurements of phosphate and arsenate were made with a Varian Series 634 (double-beam) spectrophotometer, equipped with a Varian Recorder Model 9176, and fitted with a specially designed thermostated bath system keeping the reaction conditions in the 1-cm cuvette at a pre-selected temperature. Temperature-acclimatized water samples and reagents were mixed in Pyrex glass tubes and introduced to the cuvette within 30 sec. Other arsenate and phosphate determinations were conducted using ordinary techniques, i.e. simply mixing the sample solutions with reagents in Pyrex glass

tubes and light absorbance recording by a Philips PU 8740 UV/Vis (single-beam) spectrophotometer, equipped with a Philips Plotter Printer, and carried out at room temperature (+20°C). Likewise, the solutions with fluoride and silicate were prepared in Pyrex glass tubes, but the colorimetric measurements were conducted with a Hitachi U-2000 (doublebeam) spectrophotometer. Light absorbance was recorded at 880 nm throughout.

RESULTS AND DISCUSSION

The procedural modifications of Murphy and Riley,¹ Koroleff² and Parsons *et al.*³ differ with respect to the recommended reagent concentration of molybdenum (5.4, 2.7 and 3.1 mM, respectively), antimony (0.068, 0.059 and 0.038 mM, respectively), ascorbic acid (4.80, 10.92 and 5.57 mM, respectively) and sulphuric acid (0.20, 0.10, 0.11 M, respectively) in the final solution. The effect of acidity on the reaction is complex, since it influences the rate as well as the amount and speciation of complexes formed.^{14,24–28} We therefore compared the three procedures with respect to the time needed to yield maximum absorbance of phosphoantimonylmolybdenum blue and followed its stability by continuous time course recording. However, within the concentration range of 50–500 µg PO₄-P L⁻¹, no appreciable differences were found between the three procedures; all reached a constant absorbance level within 5 min (Figure 1a), and did not fade significantly during 1.5 h. A maximum absorbance of phosphoantimonylmolybdenum blue within 5 min during normal room temperature



Figure 1 a Time course of formation of the phosphoantimonylmolybdenum blue complex, following procedures of Murphy and Riley¹ (M&R), Koroleft² (K) and Parsons *et al.*³ (P), in water containing phosphate of a concentration of 50 and 500 μ g PO₄-P L⁻¹, respectively. Reaction temperature: +20°C. **b** Time course of formation of the phosphoantimonylmolybdenum blue complex (according to Koroleff²) at three different reaction temperatures, +15°, +20° and +25°C of water containing phosphate in a concentration of 50, 100 and 500 μ g PO₄-P L⁻¹, respectively.

conditions accords with what is previously reported for a more narrow phosphate concentration range of 155–250 μ g PO₄-P L⁻¹.^{13–14} Thus, we restricted further evaluation to the modification of Koroleff.²

We compared the time taken to reach maximum absorbance level, at three different temperatures, $+15^{\circ}$, $+20^{\circ}$ and $+25^{\circ}$ C, and three different phosphate concentrations, 50, 100 and 500 µg PO₄-P L⁻¹. As shown in Figure 1b, within the temperature range studied (roughly normal room temperature range), only slight differences were found. Colour development tended to be somewhat slower at lower temperatures and in solutions of lower phosphate concentrations. Caution is advised if measurements are made at temperatures below $+15^{\circ}$ C, since the time necessary to reach maximum absorbance may then be longer than shown in Figure 1b.¹³

Arsenate forms a blue molybdate complex, which absorbs light at 880 nm. However, the reaction between arsenate and molybdate (Figure 2) is slower than between phosphate and molybdate (Figure 1). For arsenate concentrations lower than considered in the present study, even longer reaction times are needed for reaching maximum absorbance.^{12,26} Once maximum colour development is attained, the arsenate complex may increase the apparent absorption value considerably. Based on relations in Figure 3, the absorbance of 100 μ g AsO₄-As L⁻¹ corresponds to 74 μ g PO₄-P L⁻¹, or an optical density ratio of the arsenate to phosphate complexes of 1.35. This is lower than a ratio of 2.6 in distilled water, calculated from data in the original paper of Murphy and Riley^{1, p.35} and a ratio of about 2 in seawater, revealed from a graph presented by Jones.^{12, p.20} One way to reduce the over-estimation of phosphate due to arsenate interference is to time the reading to the shorter reaction time needed for formation of the phosphoantimonylmolybdenum complex. This implies measurement after about 5 min (Figure 1), but is useful only at low or moderate arsenate concentrations are present (see below).



Figure 2 Time course of formation of the arsenomolybdenum blue complex (according to Koroleff²), at three different reaction temperatures, $+15^{\circ}$, $+20^{\circ}$ and $+25^{\circ}$ C, in water containing arsenate in a concentration of 0.2 and 1.0 mg AsO4-As L⁻¹, respectively.



Figure 3 Apparent light absorbance as a function of varying arsenate concentration, ranging 0–1 mg AsO₄-As L^{-1} , in water solutions with a constant phosphate concentration of 50 µg PO₄-P L^{-1} (according to Koroleff²). Linear regression equation: $y = 0.460 \times +0.031$ (Pearson product-moment correlation coefficient r = 0.999, n = 5, p < 0.001). The intercept value refers to light absorbance of phosphate alone. Measurements performed at +20°C.

Arsenic typically occurs in concentrations of a few μ g L⁻¹, or less, in lakes,²⁹⁻³¹ rivers,^{29-30,32-33} coastal waters^{32,34-40} and oceans.⁴¹⁻⁴³ This implies significant arsenate interference with phosphate estimates only at low phosphate concentrations. However, arsenic concentrations of over 10 μ g As L⁻¹, and sometimes even higher than those studied here (Figures 2 and 3), have been reported from anthropogenically polluted areas, such as waters near mines, metal works and related activities,^{35,44-51} in supernatant water of sediment incubation experiments,⁵²⁻⁵³ in sediment pore water,^{50,54-55} in lakes loaded with herbicide,⁵⁶⁻⁵⁹ and also from some groundwater wells,^{46,59-60} lakes,⁶¹ streams and rivers,^{29,37,42,45} and in waters influenced by geothermal activity.⁶¹⁻⁶⁶ As a rule, it seems prudent to keep in mind the risk of interference from arsenate when measuring phosphate by the molybdenum blue method.



Figure 4 Time course for the formation of phosphoantimonylmolybdenum blue complex (according to Koroleff²) in water solutions containing three different phosphate concentrations, 50, 250 and 500 μ g PO₄-P L⁻¹, respectively, and varying concentrations of fluoride (0, 2, 4, 20, 40, 100, 150 and 200 mg F L⁻¹). Measurements performed at +20°C.

The interference of fluoride contrasts to that of arsenate, since fluoride does not affect the maximum absorbance level—in other words, causes no additional blue tint, nor does it diminish the final colour intensity. However, high fluoride concentrations give rise to slower colour development of phosphoantimonylmolybdenum blue (Figure 4). Regarding Figure 4, inhibition effects of fluoride are recorded for concentrations of at least 20 mg F L⁻¹, with an enhanced effect as fluoride concentrations increases. For a solution containing 200 mg F L⁻¹, several hours are needed for complete colour development. However, if the fluoride concentration is not too high (\leq 40 mg F L⁻¹) and the concentration of phosphate is not too low (\geq 50 µg PO₄-P L⁻¹) a reaction time of 5 min is still sufficient for complete colour development. Further reaction rate studies on waters of phosphate concentrations below 50 μ g PO₄-P L⁻¹ are desirable.

The actual mechanism behind the fluoride inhibition remains unclear. Wavelength spectra recorded while the phosphoantimonylmolybdenum complex was being formed, with and without fluoride, as well as after full colour development, were very similar, and did not deviate from those previously reported^{1,13,67} for the completed complex.²⁷ However, fluoride is known as a small, highly electronegative and reactive ion which readily replaces oxygen and hydroxyl groups, implying several different possible interference reactions from fluoride in the phosphoantimonylmolybdenum blue procedure, i.a. with the molybdate reagent⁶⁸ and phosphate itself.^{69–70} Easy ways to reduce interference from high fluoride concentrations are simply to dilute the sample or postpone the photometric recording according to the time needed to reach the maximum absorbance level (Figure 4).

Fluoride is a major constituent of seawater, usually found in concentrations of about 1–2 mg F L⁻¹ in ocean water,⁷¹⁻⁷² and in 10-fold lower concentration in rivers and lakes,⁷³⁻⁷⁴ with concentration in esturine waters being intermediate.⁷⁴⁻⁷⁶ However, concentrations above 20 mg F L⁻¹ are reported from saline lakes,⁷⁷ groundwater wells,⁷⁸⁻⁷⁹ thermal springs,^{64,80} soil extracts⁸¹ and industrially polluted river water.⁸² Even much higher fluoride concentrations (>200 mg F L⁻¹) have been reported from certain soda lakes⁷⁷ and underground brines,⁷⁸ from acid-spring waters in active volcanic areas⁸³ and in heavily polluted waters.⁸⁴⁻⁸⁵

Silicate provides an example of a component complicating the evaluation of interfering factors in the phosphate determination. In natural waters, dissolved silica occurs predominantly as protonated monomeric silicate—in other words, undissociated orthosilicic acid H₄SiO₄.^{86–88} In the present study, when the reagents for phosphate analysis with the phosphoantimonylmolybdenum blue method were simply mixed with dissolved silicate, a faint, linearly and slowly formed blue tint was recorded at 880 nm. Measurements at 50 mg SiO₄-Si L⁻¹ resulted, after 10 min of reaction, in an apparent light absorbance value corresponding to about 2 µg PO₄-P L⁻¹. After 30 min, the corresponding figure was about 6 µg PO₄-P L⁻¹, which is roughly similar to the absorbance value given by Koroleff.² However, this is considerably less than reported by Noriki,⁸⁹ and at +70°C by Pai *et al.*¹⁴ In the study of Noriki,⁸⁹ it appears that self-reduction of molybdate ion due to a too low sulphuric acid concentration has been misinterpreted as absorbance by silicate.^{14,27}

If silicate is added in combination with fluoride in the phosphate determination, the inhibition effect from fluoride is mitigated, resulting in less time needed to reach the maximum absorbance level of the phosphoantimonylmolybdenum blue complex (Figure 5). For a constant weight ratio of fluoride to silicon of four, as shown in Figure 6, the counteraction of silicate on fluoride interference is most pronounced when the fluoride concentration is extremely high (200 mg F L⁻¹). Conversely, high fluoride concentrations reduce or totally inhibit the development of the blue complex between silicate and molyb-date. These findings demonstrate interaction between silicate and fluoride, probably due to equilibrium complexation.⁶⁹

Dissolved silicate concentrations ($\geq 5 \text{ mg SiO}_4$ -Si L⁻¹) as high as those we have found to mitigate fluoride inhibition of phosphoantimonylmolybdenum blue have been reported in water from lakes,^{77,90-96} surface drain water⁹⁷ and groundwater samples,^{78,98-99} warm and cold springs,^{64,96,99} streams, rivers and the low salinity zone of some estuaries.^{75,79,90,93,98-100} In the



Figure 5 Time course for the formation of phosphoantimonylmolybdenum blue complex (according to Koroleff²) in water solutions containing constantly 250 μ g PO₄-P L⁻¹ and 100 and mg F L⁻¹, but varying concentrations of silicate: 0, 5, 10, 20, 40 and 100 mg SiO₄-Si L⁻¹, respectively. Measurements performed at +20°C.

true marine realm, such high silicate concentrations are not commonly found, ¹⁰¹⁻¹⁰³ but have been reported from deep waters of the Bering Sea, ¹⁰⁴⁻¹⁰⁵ the Sea of Okhotsk¹⁰⁴ and the northern Pacific, ¹⁰⁴⁻¹⁰⁵ from hydrothermal deep-sea vents, ^{62,106-108} deep brine of the Red Sea, ¹⁰⁹ and euxinic water of the Black Sea, ¹¹⁰⁻¹¹¹ in supernatant water of sediment incubation experiments¹¹²⁻¹¹³ and in sediment pore waters. ¹¹⁴⁻¹¹⁸

Retardation of the colour development rate of the phosphomolybdenum blue complex is an interference effect previously reported for stannic ion, tungstate ion¹¹⁹ and vanadate.¹²⁰ The latter two oxoanions are known to form heteropolyanions with phosphate as well as molybdate,¹²¹⁻¹²² suggesting competitive interaction with the phosphomolybdenum blue complex formation as a possible causal explanation of slow colour development. This can be expected to be valid also for stannic ion.¹²³ Retardation effects such as are found for fluoride (Figure 4), or relationships describing the counteracting effect from silicate on the fluoride interference (Figures 5 and 6), seem not to have been reported before. However, already Denigès²⁰ noted that interference of fluoride interference appears to be valid for borate^{20,22-23} and a slowed phosphomolybdenum complex formation, analogous to what is found for fluoride, may be caused by hypochlorite or chlorine gas.¹²⁴ Element interactions resulting in retarded colour development provide an analytical source of error difficult to recognize, which may cause under-estimation of the true phosphate concentration. Further studies on this type of interference effects seem justified.



Figure 6 Time course for the formation of phosphoantimonylmolybdenum blue complex (according to Koroleff²) in water solutions containing three different phosphate concentrations, 50, 250 and 500 μ g PO₄-P L⁻¹, respectively, and varying concentrations of fluoride (0, 8, 20, 40, 100 and 200 mg F L⁻¹) and silicate (0, 2, 5, 10, 25 and 50 mg SiO₄-Si L⁻¹), at a constant element weight ratio of four. Measurements performed at +20°C.

CONCLUSIONS

In the studied range of 50–500 µg PO₄-P L⁻¹, analytical procedures according to Murphy and Riley,¹ Koroleff² and Parsons *et al.*³ all resulted in complete colour development of the phosphoantimonylmolybdenum blue complex within 5 min after reagent addition. Reducing the reaction temperature from +25° to +15°C slowed the formation rate of the blue complex, but not enough to be of practical importance. Arsenate, fluoride and silicate interfere in the analysis of phosphate in different ways.

Arsenate forms a blue complex with molybdate, albeit at a slower rate than phosphate. This interference of arsenate may cause elevated absorption values and over-estimation of the true phosphate concentration. Of course, the potential bias of this interference is highest for waters with low phosphate concentrations^{56–57,125} or in waters with high arsenate to phosphate ratios.⁵⁶ The slow formation of the arsenate complex means that the interference can be minimized by recording the absorbance value as soon as the phosphoantimonylmolybdenum complex has developed fully. This effectively reduces the interference of arsenate on phosphate determinations in waters with low arsenate concentration. Still, high arsenate concentrations remain a potential source of error when determining phosphate by the molybdenum blue method.

High concentrations of fluoride ($\geq 20 \text{ mg F L}^{-1}$) cause slower colour development of the phosphoantimonylmolybdenum blue complex formation, but do not diminish the final colour intensity. The inhibitory effect may be caused either by competitive interaction between fluoride and phosphate for a common factor, or by a direct reaction between fluoride and phosphate.

Silicate may affect the analytical procedure in two ways. Alone, after prolonged standing, silicate can cause slightly increased light absorption, due to a blue complex formed with molybdate. High fluoride concentrations, however, reduce or totally inhibit this complexation. Conversely, silicate counteracts the inhibitory effect of fluoride in the phosphoantimonylmolybdenum complex formation, and may thereby indirectly affect the determination of phosphate.

Element interactions resulting in varying rates of colour development may be treacherous sources of analytical error. In particular, slowed formation rates of the phosphoantimonylmolybdenum complex can easily cause serious under-estimation of the true phosphate concentration. Therefore, intermittent checking of the rate of colour development, to ensure full expression, may be a useful precaution in spectrophotometric analysis, particularly when new types of samples are analyzed. An easy way to reduce interference by arsenate, fluoride as well as silicate is to simply dilute the sample prior to addition of reagents, provided that phosphate concentrations are sufficiently high.

Acknowledgements

We are grateful to Ragnar Elmgren and Anneli Gunnars for reviewing the manuscript, and to Birgitta Holmstedt and Bodil Kajrup for help with the chemical analyses. This study was supported by the Swedish Environmental Protection Agency.

References

- 1. J. Murphy and J. P. Riley, Anal. Chim. Acta, 27, 31-36 (1962).
- F. Koroleff, in: *Methods of Seawater Analysis* (K. Grasshoff, M. Ehrhardt and K. Kremling, eds.), Verlag Chemie, Weinheim, 1983, 2nd ed., pp. 125–139.
- 3. T. R. Parsons, Y. Maita and C. M. Lalli, A Manual of Chemical and Biological Methods for Seawater Analysis, Pergamon Press, Oxford, 1984.
- 4. M. F. Osmond, Bull. Soc. Chim. Paris, 47, 745-748 (1887).

- 5. F. A. J. Armstrong, Oceanogr. Mar. Biol. Ann. Rev., 3, 79-93 (1965).
- J. P. Riley, in: Chemical Oceanography Vol. 2, (J. P. Riley and G. Skirrow, eds., Academic Press, London, 1965) pp. 295–424.
- S. Olsen, in: Chemical Environment in the Aquatic Habitat (H. L. Golterman and R. S. Clymo, eds., N. V. Noord-Hollandsche Uitgevers Maatschappij, Amsterdam, 1967) pp. 63–105.
- 8. J. D. Burton, Wat. Res., 7, 291-307 (1973).
- J. E. Harwood and W. H. J. Hattingh, in: Environmental Phosphorus Handbook (E. J. Griffith, A. Beeton, J. M. Spencer and D. T. Mitchell, eds., Wiley & Sons, New York, 1973) pp. 289–301.
- R. A. Kimerle and W. Rorie, in: *Environmental Phosphorus Handbook* (E. J. Griffith, A. Beeton, J. M. Spencer and D. T. Mitchell, eds., Wiley & Sons, New York, 1973) pp. 367–379.
- O. G. Koch and G. A. Koch-Dedic, Handbuch der Spurenanalyse, (Springer-Verlag Berlin, 1974) 2nd ed., pp. 895–921.
- 12. P. G. W. Jones, J. Mar. Biol. Ass. U.K., 46, 19-32 (1966).
- 13. Y. S. To and C. W. Randall, J. Wat. Pollut. Contr. Fed., 49, 689-692 (1977).
- 14. S.-C. Pai, C.-C. Yang and J. P. Riley, Anal. Chim. Acta, 229, 115-120 (1990).
- J. D. H. Strickland and T. R. Parsons, A manual of sea water analysis (Bull. Fish. Res. Bd Can. 125, 1965), 2nd ed.
- F. Koroleff, in: New Baltic Manual, with Methods for Sampling and Analyses of Physical, Chemical and Biological Parameters. (S. R. Carlberg, ed., Cooperative Research Report Ser. A, No. 29, International Council for the Exploration of the Sea, Charlottenlund, Denmark, 1972) pp. 44–49.
- J. D. H. Strickland and T. R. Parsons, A Practical Handbook of Seawater Analysis (Bull. Fish. Res. Bd Can. 167, 1972), 2nd ed.
- H. L. Golterman, R. S. Clymo and M. A. M. Ohnstad, *Methods for Physical and Chemical Analysis of Fresh Waters* (IBP Handbook No. 8, Blackwell Scientific Publications, Oxford, 1978) 2nd ed.
- 19. R. G. Wetzel and G. E. Likens, Limnological Analyses (Springer-Verlag, New York, 1991) 2nd ed.
- 20. G. Denigès, C. R. Hebd. Séanc. Acad. Sci., Paris, 171, 802-804 (1920).
- 21. D. Florentin, Ann. Chim. Anal. Chim. Appl. Rev. Chim. Anal., 3, 295-296 (1921).
- 22. M. v. Wrangell and W. Stollenwerk, Landw. Jahrb., 63, 674-676 (1926).
- 23. J. Tischer, Z. Pflanzenernähr. Düng. Bodenk., A 33, 192-242 (1934).
- 24. S. R. Crouch and H. V. Malmstadt, Anal. Chem., 39, 1084-1089 (1967).
- 25. A. C. Javier, S. R. Crouch and H. V. Malmstadt, Anal. Chem., 40, 1922-1925 (1968).
- 26. D. L. Johnson and M. E. Q. Pilson, Anal. Chim. Acta, 58, 289-299 (1972).
- 27. J. E. Going and S. J. Eisenreich, Anal. Chim. Acta, 70, 95-106 (1974).
- 28. L. Pettersson, I. Andersson and L.-Ö. Öhman, Inorg. Chem., 25, 4726-4733 (1986).
- 29. M. O. Andreae, Deep-Sea Res., 25, 391-402 (1978).
- U. Förstner and G. T. W. Wittmann, Metal Pollution in the Aquatic Environment (Springer-Verlag, Berlin, 1981) 2nd ed.
- G. Lithner, Bedömningsgrunder för sjöar och vattendrag. Bakgrundsdokument 2. Metaller (Swedish Environmental Protection Agency Rept. 3628, Solna, Sweden, 1989).
- R. Carpenter, M. L. Peterson and R. A. Jahnke, in: *Estuarine Interactions* (M. L. Wiley, ed., Academic Press, New York, 1978) pp. 459–480.
- 33. D. G. Waslenchuk, Chem. Geol., 24, 315-325 (1979).
- 34. M. O. Andreae, J. T. Byrd and P. N. Froelich, Jr., Environ. Sci. Technol., 17, 731-737 (1983).
- 35. W. J. Langston, Can. J. Fish. Aquat. Sci., 40 (Suppl. 2), 143-150 (1983).
- 36. A. G. Howard, M. H. Arbab-Zavar and S. Apte, Est. Coast. Shelf Sci., 19, 493-504 (1984).
- H. A. van der Sloot, D. Hoede, J. Wijkstra, J. C. Duinker and R. F. Nolting, *Est. Coast. Shelf Sci.*, 21, 633–651 (1985).
- 38. M. Stoeppler, M. Burow, F. Backhaus, W. Schramm and H. W. Nürnberg, Mar. Chem., 18, 321-334 (1986).
- 39. J. T. Byrd, Mar. Chem., 25, 383-394 (1988).
- 40. G.-H. Tremblay and C. Gobeil, Mar. Pollut. Bull., 21, 465-469 (1990).
- 41. K. Sugawara, K. Terada, S. Kanamori, N. Kanamori and S. Okabe, J. Earth Sci., Nagoya Univ., 10, 34–50 (1962).
- 42. H. A. van der Sloot, D. Hoede and J. Wijkstra, Netherl. J. Sea Res., 23, 379-386 (1989).
- J. D. Burton and P. J. Stratham, in: *Heavy Metals in the Marine Environment* (R. W. Furness and P. S. Rainbow, eds., CRC Press, Boca Raton, FL, 1990) pp. 5–25.
- G. Lithner, Rönnskärsutredningen 1973 (National Swedish Environment Protection Board, PM 497, Solna, Sweden, 1974).

- 45. S. R. Aston, I. Thornton, J. S. Webb, B. L. Milford and J. B. Purves, Sci. Tot. Environ., 4, 347-358 (1975).
- 46. D. A. Grantham and J. F. Jones, J. Am. Wat. Wks Ass., 69, 653-657 (1977).
- R. Wagemann, N. B. Snow, D. M. Rosenberg and A. Lutz, Arch. Environ. Contam. Toxicol., 7, 169–191 (1978).
- 48. D. W. Klumpp and P. J. Peterson, Environ. Pollut., 19, 11-20 (1979).
- R. R. Brooks, J. E. Fergusson, J. Holzbecher, D. E. Ryan, H. F. Zhang, J. M. Dale and B. Freedman, *Environ*. *Pollut.*, **B 4**, 109–117 (1982).
- 50. A. G. Howard, S. C. Apte, S. D. W. Comber and R. J. Morris, Est. Coast. Shelf Sci., 27, 427-443 (1988).
- 51. M. D. Zingde, S. Y. S. Singbal, C. F. Moraes and C. V. G. Reddy, Indian J. Mar. Sci., 5, 212-217 (1976).
- 52. M. Edgren and M. Notter, *The flux of metals from contaminated bottoms of the Skellefte Inlet* (in Swedish with English summary) (National Swedish Environment Protection Board, PM **1081**, Solna, Sweden, 1979).
- 53. N. G. Holm, Chem. Geol., 68, 89-98 (1988).
- 54. M. L. Peterson and R. Carpenter, Geochim. Cosmochim. Acta, 50, 353-369 (1986).
- 55. J. Cornett, L. Chant and B. Risto, Hydrobiologia, 235/236, 533-544 (1992).
- 56. W. Chamberlain and J. Shapiro, Limnol. Oceanogr., 14, 921-927 (1969).
- 57. J. Shapiro, Science, 171, 234 (1971).
- 58. S. A. Lis and P. K. Hopke, Environ. Lett., 5, 45-51 (1973).
- 59. R. E. Foley, J. R. Spotila, J. P. Giesy and C. H. Wall, Environ. Biol. Fish., 3, 361-367 (1978).
- W. P. Tseng, H. M. Chu, S. W. How, J. M. Fong, C. S. Lin, and S. Yeh, J. Nat. Cancer Inst., 40, 453–463 (1968).
- H. Onishi, in: Handbook of Geochemistry Vol II/3 (K. H. Wedepohl, ed., Springer-Verlag, Berlin, 1969) pp. 33-I-1-33-I-6.
- 62. K. L. Von Damm, Ann. Rev. Earth Planet. Sci., 18, 173-204 (1990).
- 63. P. F. Reay, J. Appl. Ecol., 9, 557-565 (1972).
- 64. J. J. Rowe, R. O. Fournier and G. W. Morey, U.S. Geol. Surv. Bull., 1303, 1-31 (1973).
- 65. C. Cremisini, M. Dall'Aligo and E. Ghiara, in: International Conference, Management and Control of Heavy Metals in the Environment (CEP Consultants Edinburgh, 1979) pp. 341–344.
- 66. J. Aggett and G. A. O'Brien, Environ. Sci. Technol., 19, 231-238 (1985).
- 67. D. L. Johnson, Environ. Sci. Technol., 5, 411-414 (1971).
- 68. A. G. Sharpe, in: Fluorine Chemistry Vol. 2, (J. H. Simons, ed., Academic Press, New York, 1954) pp. 1-37.
- 69. W. Lange, in: Fluorine Chemistry Vol. 1, (J. H. Simons, ed., Academic Press, New York, 1950) pp. 125-188.
- R. Schmutzler, in: Advances in Fluorine Chemistry Vol. 5, (M. Stacey, J. C. Tatlow and A. G. Sharp, eds., Butterworths, London, 1965) pp. 31-285.
- F. Culkin, in: Chemical Oceanography Vol. 1, (J. P. Riley and G. Skirrow, eds., Academic Press, London, 1965) pp. 121-161.
- T. R. S. Wilson, in: Chemical Oceanography Vol. 1 (J. P. Riley and G. Skirrow, eds., Academic Press, London, 1975) 2nd ed., pp. 365–413.
- 73. S. W. Reeder, B. Hitchon and A. A. Levinson, Geochim. Cosmochim. Acta, 36, 825-865 (1972).
- 74. B. Kullenberg and R. Sen Gupta, Geochim. Cosmochim. Acta, 37, 1327-1337 (1973).
- P. S. Liss, in: Estuarine Chemistry (J. D. Burton and P. S. Liss, eds., Academic Press, London, 1976) pp. 93-130.
- S. N. de Sousa and R. Sen Gupta, Indian J. Mar. Sci., 17, 195–201 (1988).
- 77. P. Kilham and R. E. Hecky, Limnol. Oceanogr., 18, 932-945 (1973).
- 78. G. W. Bond, Mem. Geol. Surv. S. Afr., 41, 1-208 (1946).
- J. D. Hem, Quality of water of the Gila River Basin above Coolidge Dam, Arizona (Geological Survey Water-Supply Paper No. 1104, U.S. Government Printing Office, Washington, DC, 1950).
- K. Sugawara 1954, cited by C. W. Correns p. 205, in: *Physics and Chemistry of the Earth* Vol. 1, (L. A. Ahrens, K. Rankama and S. K. Runcorn, eds., Pergamon Press, London, 1956) pp. 181-233.
- 81. K. C. Walton, Sci. Tot. Environ., 65, 247-256 (1987).
- 82. D. J. Moore, Chesapeake Sci., 12, 1-13 (1971).
- 83. W. A. J. Mahon, New Zeal. J. Sci., 7, 3-28 (1964).
- 84. L. Karstad, Bull. Wildl. Dis. Ass., 3, 42-46 (1967).
- 85. B. S. Krumgalz, G. Fainshtein, L. Gorfunkel and Y. Nathan, Est. Coast. Shelf Sci., 30, 1-15 (1990).
- 86. K. B. Krauskopf, Geochim. Cosmochim. Acta, 10, 1-26 (1956).
- 87. J. D. Burton, T. M. Leatherland and P. S. Liss, Limnol. Oceanogr., 15, 473-476 (1970).
- N. Ingri, in: Biochemistry of Silicon and Related Problems (G. Bendz and I. Lindquist, eds., Plenum Press, New York, 1978) pp. 3-51.

- 89. S. Noriki, J. Oceanogr. Soc. Jap., 39, 324-326 (1983).
- 90. D. A. Livingstone, *Chemical composition of rivers and lakes.* (Data of Geochemistry, Geological Survey Professional Paper No. **440-G**, U.S. Government Printing Office, Washington, DC, 1963) 6th ed.
- 91. J. F. Talling and I. B. Talling, Int. Revue ges. Hydrobiol., 50, 421-463 (1965).
- 92. U. Tessenow, Arch. Hydrobiol., Suppl. 32, 1-136 (1966).
- 93. O. Amit, Chem. Geol., 5, 121-129 (1969).
- H. P. Eugster and L. A. Hardie, in: Lakes: Chemistry, Geology, Physics (A. Lerman, ed., Springer-Verlag, New York, 1978) pp. 237-293.
- 95. R. G. Wetzel, Limnology (Saunders College Publ. Philadelphia, PA, 1983) 2nd ed.
- 96. R. B. Wood and J. F. Talling, Hydrobiologia, 158, 29-67 (1988).
- 97. F. M. Eaton, G. W. McLean, G. S. Bredell and H. E. Doner, Soil Sci., 105, 260-280 (1968).
- 98. S. N. Davis, Am. J. Sci., 262, 870-891 (1964).
- 99. D. E. White, W. W. Brannock and K. J. Murata, Geochim. Cosmochim. Acta, 10, 27-59 (1956).
- J. Kobayashi, in: Chemical Environment in the Aquatic Habitat (H. L. Golterman and R. S. Clymo, eds. N. V. Noord-Hollandsche Uitgevers Maatschappij, Amsterdam, 1967) pp. 41–55.
- F. A. J. Armstrong, in: Chemical Oceanography Vol. 1 (J. P. Riley and G. Skirrow, eds., Academic Press, London, 1965) pp. 409–432.
- C. P. Spencer, in: Chemical Oceanography Vol. 2 (J. P. Riley and G. Skirrow, eds., Academic Press, London, 1975) 2nd ed., pp. 245-300.
- C. P. Spencer, in: Silicon Geochemistry and Biogeochemistry (S. R. Aston, ed., Academic Press, London, 1983) pp. 101-141.
- 104. J. M. Edmond, S. S. Jacobs, A. L. Gordon, A. W. Mantyla and R. F. Weiss, J. Geophys. Res., 84, 7809–7826 (1979).
- 105. S. Tsunogai, M. Kusakabe, H. Iizumi, I. Koike, and A. Hattori, Deep-Sea Res., 26, 641-659 (1979).
- J. B. Corliss, J. Dymond, L. I. Gordon, J. M. Edmond, R. P. von Herzen, R. D. Ballard, K. Green, D. Williams, A. Bainbridge, K. Crane and T. H. van Andel, *Science*, 203, 1073–1083 (1979).
- 107. D. M. Karl, G. M. McMurtry, A. Malahoff and M. O. Garcia, Nature, 335, 532-535 (1988).
- 108. D. M. Karl, A. M. Brittain, and B. D. Tilbrook, Deep-Sea Res., 36, 1655-1673 (1989).
- P. G. Brewer and D. W. Spencer, in: Hot Brines and Recent Heavy Metal Deposits in the Red Sea (E. T. Degens and D. A. Ross, eds., Springer-Verlag, New York, 1969) pp. 174–179.
- K. Grasshoff, in: Chemical Oceanography Vol. 2 (J. P. Riley and G. Skirrow, eds., Academic Press, London, 1975) 2nd ed., pp. 455–597.
- Y. I. Sorokin, in: *Estuaries and Enclosed Seas* (B. H. Ketchum, ed., Ecosystems of the World 26, Elsevier, Amsterdam, 1983) pp. 253-292.
- 112. C. H. Mortimer, J. Ecol., 29, 280-329 (1941).
- 113. A. Gunnars, Exchange of phosphorus and silicon over the sediment-water interface during positive redox-turnover—the role of iron and manganese, Chem. Comm. 4 (Dept. Phys. Inorg. Struct. Chem., Univ. Stockholm, Sweden, 1990).
- 114. R. Siever, K. C. Beck and R. A. Berner, J. Geol., 73, 39-73 (1965).
- 115. D. R. Schink, K. A. Fanning and M. E. Q. Pilson, J. Geophys. Res., 79, 2243-2250 (1974).
- F. T. Manheim, in: Chemical Oceanography Vol. 6 (J. P. Riley and R. Chester, eds., Academic Press, London, 1976, 2nd ed.) pp. 115–186.
- H. Elderfield, R. J. McCaffrey, N. Luedtke, M. Bender and V. W. Truesdale, Am. J. Sci., 281, 1021-1055 (1981).
- 118. J. E. Mackin, Am. J. Sci., 287, 197-241 (1987).
- 119. J. C. van Schouwenburg, and I. Walinga, Anal. Chim. Acta, 37, 271-274 (1967).
- 120. D. F. Boltz and M. G. Mellon, Indust. Eng. Chem., Anal. Ed., 19, 873-877 (1947).
- D. L. Kepert, in: Comprehensive Inorganic Chemistry Vol. 4, (J. C. Bailar, Jr., H. J. Emeléus, R. Nyholm and A. F. Trotman-Dickenson, eds., Pergamon Press, Oxford, 1973) pp. 607–672.
- 122. M. T. Pope, Heteropoly and Isopoly Oxometalates, (Springer-Verlag, Berlin, 1983).
- 123. A. Schouten and B. Cros, Polyhedron, 1, 283-287 (1982).
- 124. I. Dellien and L.-Å. Johansson, Vatten, 37, 349–353 (1981).
- 125. D. L. Johnson and M. E. Q. Pilson, J. Mar. Res., 30, 140-149 (1972).